

Sodium Azide									
Approximate Cell Density x 10 ⁸ /ml	1.5	3	6	9	12	15	18	21	24
Absorbance Specification at 625nm (ABS)	0.08 to 0.10	0.14 to 0.17	0.27 to 0.31	0.38 to 0.42	0.51 to 0.55	0.67 to 0.70	0.74 to 0.77	0.83 to 0.88	0.94 to 0.98

STORAGE AND SHELF LIFE

Storage: Upon receipt store at 2-30°C. These products are ready for use and no further preparation is necessary. These products should be stored in their original container. Do not freeze or overheat. Do not incubate prior to use. Standards should not be used if there are any signs of deterioration, contamination, color change or if the expiration date has passed.

The expiration date on the product label applies to the product in its intact packaging when stored as directed. The product may be used and tested up to the expiration date on the product label and incubated for the recommended incubation times as stated below.

Refer to the document "[Storage](#)" for more information.

PRECAUTIONS

This product may contain components of animal origin. Certified knowledge of the origin and/or sanitary state of the animals does not guarantee the absence of transmissible pathogenic agents. Therefore, it is recommended that these products be treated as potentially infectious, and handle observing the usual Universal Precautions for blood. Do not ingest, inhale, or allow to come into contact with skin.

This product is for *in vitro* diagnostic use only. It is to be used only by adequately trained and qualified laboratory personnel. Observe approved biohazard precautions and aseptic techniques. All laboratory specimens should be considered infectious and handled according to "standard precautions." Refer to the document "[Guidelines for Isolation Precautions](#)" from the Centers for Disease Control and Prevention.

For additional information regarding specific precautions for the prevention of the transmission of all infectious agents from laboratory instruments and materials, and for recommendations for the management of exposure to infectious disease, refer to CLSI document M29: *Protection of Laboratory Workers from Occupationally Acquired Infections*.

Sterilize all biohazard waste before disposal.

Refer to the document "[Precautions When Using Media](#)" for more information.

PROCEDURE

Specimen Collection: Not applicable since these standards are used for the standardization of inoculum for susceptibility testing. Isolation techniques and tests for purity are necessary. Information on specimen collection may be found in standard reference texts.

Method of Use:

1. Mix the standard to fully suspend the particles. Gently mix the McFarland Latex Standards (ML05-ML8) by inverting - **Do not vortex** the latex standards.
2. Adjust the turbidity of an actively growing broth culture or bacterial suspension of colonies selected from an 18-24 hour agar plate to obtain a turbidity visually comparable to that of the desired standard. The tubes for the suspension should be the same diameter as the McFarland Latex Standard tube.
3. For visual comparison, use adequate light and read the tubes against a white card with contrasting black lines (Wickerham Card, Cat. no. Z08).⁽³⁾

INTERPRETATION OF RESULTS

Equal obliteration or distortion of contrasting black lines on the white background indicates a turbidity match.

LIMITATIONS

Do not use a mechanical mixer (vortex) with latex standards.

Broth media that is dark yellow, orange or brown will not provide the proper contrast with McFarland Latex Standards, possibly resulting in bacterial suspensions of incorrect densities. Trial comparisons should be performed.⁽⁵⁾ Use adequate light and read the standard and test broth against a white card with contrasting black lines (Wickerham Card, Cat. no. Z08).⁽³⁾

The tubes for the suspension should be the same diameter as the McFarland Latex Standard tube.

Bacterial densities may be too heavy when colonies of *Haemophilus influenzae* < 24 hours old are used to prepare suspensions.⁽⁶⁾

McFarland Standards are recommended when performing visual comparisons or when using a spectrophotometer adjusted to the proper setting.^(5,7) Use with instruments which use alternative light sources, such as scattered light, have not been established.

Refer to the document "[Limitations of Procedures and Warranty](#)" for more information.

MATERIALS REQUIRED BUT NOT PROVIDED

Standard microbiological supplies and equipment such as loops, broth media, vortex mixer and Wickerham Card (Cat. no. Z08), etc., are not provided.

QUALITY CONTROL

All lot numbers of McFarland Latex Standards are tested spectrophotometrically and have been found to be acceptable.

PHYSICAL APPEARANCE

McFarland Latex Standards appear white with a bluish tinge.



McFarland Latex Standards

REFERENCES

1. Versalovic, J., et al. *Manual of Clinical Microbiology*. American Society for Microbiology, Washington, D.C.
2. Tille, P., et al. *Bailey and Scott's Diagnostic Microbiology*, C.V. Mosby Company, St. Louis, MO.
3. Clinical and Laboratory Standards Institute (CLSI - formerly NCCLS). 2009. *Performance Standards for Antimicrobial Disk Susceptibility Tests*; Approved Standard - Tenth Edition. M02-A10. CLSI, Wayne, PA.
4. McFarland, J. 1907. Nephelometer: An Instrument for Estimating the Number of Bacteria in Suspension Used for Calculating the Opsonic Index for Vaccines. *Journ. American Med. Assoc.*;14: 1176-1178.
5. Perry, J.L. and G.R. Miller. 1992. Abstract, ASM.
6. Doern, G.V. and R.N. Jones. 1988. Antimicrobial Susceptibility Testing of *Haemophilus influenzae*, *Branhamella catarrhalis* and *Neisseria gonorrhoeae*. *Antimicrob. Agents and Chemothr.*; 32: 1747-1753.
7. Carlberg, D.M. Determining the Effect of Antibiotics on Bacterial Growth by Optical and Electrical Methods in *Antibiotics in Laboratory Medicine*, 2nd ed. V. Lorian, editor, Chapter 3, p. 64-92, Williams & Wilkins.
8. Clinical and Laboratory Standards Institute (CLSI - formerly NCCLS). *Performance Standards for Antimicrobial Disk Susceptibility Tests*, 10th ed., M02-03, Section 8.1. CLSI, Wayne, PA.

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