



Instructions for Use

HardyCHROM™ Candida + auris

Cat. no. G343

HardyCHROM™ Candida + auris, 15x100mm Plate, 18ml

10 plates/bag

INTENDED USE

Hardy Diagnostics HardyCHROM™ Candida + auris is recommended for the selective isolation and differential identification of *Candida* species. The medium allows for the differentiation of *C. albicans*, *C. tropicalis*, *C. krusei*, and *C. auris* based on colony morphology, color, and positive UV fluorescence.

SUMMARY

Candida auris is an emerging healthcare-associated pathogen that represents a serious multi-drug resistant (MDR) global threat. Though the organism often presents fewer virulence factors than *Candida albicans*, *C. auris* may persistently colonize the skin and hospital environment, making its transmission within and between healthcare settings more difficult to control.⁽¹¹⁾ *C. auris* fungemia results in a wide range of mortality rates (from 32%-66%), depending upon the patient's overall condition, underlying disease, geographic region, access to medical care, and age.⁽¹¹⁾ A review of the organism's genome demonstrates that it harbors genes well characterized as virulence factors in other *Candida* species, as well as genes for biofilm production and MDR transcription factors.⁽¹¹⁾

HardyCHROM™ Candida + auris is a selective and differential medium containing chromogenic substrates. After degradation by specific enzymes, the substrates release different colored compounds. Certain species or groups of organisms can then be differentiated with a minimum number of confirmatory tests.

Colonies of *C. albicans* will appear green to dark metallic green. Colonies of *C. tropicalis* will appear medium blue to dark metallic blue with a blue halo. Colonies of *C. krusei* will appear flat, often rough or crenated, and pink to medium pink in color. Colonies of *C. auris* will appear white with a characteristic teal to teal-green "bullseye" center. Other species will appear pink, often with a darker mauve center (e.g. *C. glabrata* and other species). Other yeasts may appear white to pink. Most typical yeast colonies tested for fluorescence at 48 hours using a UV lamp at 365nm will be negative, except for *C. auris* which will be positive at 48-72 hours.

Additionally, HardyCHROM™ Candida + auris can be used in conjunction with Rapid Trehalose Broth ([Cat. no. Z205](#)) or GlabrataQuick™ ([Cat. no. Z298](#)) to aid in the identification of *C. glabrata*. When HardyCHROM™ Candida + auris is used as the primary plating medium, only colonies that morphologically (pink, often with a darker mauve center) resemble *C. glabrata* should be tested for trehalose assimilation.

HardyCHROM™ Candida + auris contains glucose and selected peptones as a nutrient supply. Chromogenic substrates are incorporated to enable the production of different colored compounds when degraded by specific enzymes formed by the yeast. Chloramphenicol is added as an inhibitory agent against the growth of most bacteria, which may be present in the sample. Agar is the solidifying agent.

FORMULA

Ingredients per liter of deionized water:*

Glucose	20.0gm
Peptone	10.0gm
Chromogenic Mixture	2.15gm
Titanium Dioxide	2.0gm
Chloramphenicol	0.5gm
Agar	15.0gm

Final pH 6.1 +/- 0.2 at 25°C.

* Adjusted and/or supplemented as required to meet performance criteria.

STORAGE AND SHELF LIFE

Storage: Upon receipt, store at 2-8°C away from direct light. Media should not be used if there are any signs of deterioration (shrinking, cracking, or discoloration), contamination, or if the expiration date has passed. Product is light and temperature sensitive; protect from light, excessive heat, moisture, and freezing.

The expiration dating on the product label applies to the product in its intact packaging when stored as directed. The product may be used and tested up to the expiration date on the product label and incubated for the recommended quality control incubation times.

Refer to the document "[Storage](#)" for more information.

PRECAUTIONS

This product may contain components of animal origin. Certified knowledge of the origin and/or sanitary state of the animals does not guarantee the absence of transmissible pathogenic agents. Therefore, it is recommended that these products be treated as potentially infectious, and handle observing the usual universal blood precautions. Do not ingest, inhale, or allow to come into contact with skin.

This product is for *in vitro* diagnostic use only. It is to be used only by adequately trained and qualified laboratory personnel. Observe approved biohazard precautions and aseptic techniques. All laboratory specimens should be considered infectious and handled according to "standard precautions." The "Guidelines for Isolation Precautions" is available from the Centers for Disease Control and Prevention at www.cdc.gov/ncidod/dhqp/gl_isolation.html.

For additional information regarding specific precautions for the prevention of the transmission of all infectious agents from laboratory instruments and materials, and for recommendations for the management of exposure to infectious disease, refer to CLSI document M-29: *Protection of Laboratory Workers from Occupationally Acquired Infections: Approved Guideline*.

Sterilize all biohazard waste before disposal.

Refer to the document "[Precautions When Using Media](#)" for more information.

Refer to the document [SDS Search](#) instructions on the Hardy Diagnostics' website for more information.

PROCEDURE

Specimen Collection: Consult listed references for information on specimen collection.⁽¹⁻⁸⁾ Infectious material should be submitted directly to the laboratory without delay and protected from excessive heat and cold. If a delay in processing is anticipated, the specimen should be inoculated onto an appropriate transport media and refrigerated until inoculation.

Consult the listed references for information regarding the processing of specimens.⁽¹⁻⁹⁾

Protect media from light during storage and incubation as the product is light sensitive.

Method of Use: Allow plates to warm to room temperature. The agar surface should be dry prior to inoculating. Inoculate and streak the specimen as soon as possible after collection. If the specimen to be cultured is on a swab, roll the swab over a small area of the agar surface. Streak for isolation with a sterile loop. Incubate plates in an inverted position, protected from the light, aerobically at 35°C. with increased humidity for 48-72 hours.*

Most pathogenic strains of yeast grow at 35°C. Some strains, other than *C. albicans*, *C. krusei*, *C. glabrata*, *C. tropicalis*, and *C. auris* may fail to grow at 35°C. If cultivation of all yeast strains is desired, the recommended incubation temperature is 30°C. for up to 7 days, since the lower temperature will slow the growth.

*In instances where *C. auris* is suspected, it is recommended to extend the incubation period to at least 72 hours. See the Limitations section for more information.

INTERPRETATION OF RESULTS

Examine plates for colonies showing typical morphology and color. Use a UV Lamp ([Cat. no. UVL56](#)) to detect colony fluorescence.

Some strains may show sufficient growth and color development when read at 24 hours; however, all plates should be incubated for at least 48 hours to allow for adequate color development and to detect fluorescence. Colors will intensify with age.

A medium size, smooth, green to dark metallic green colored colony at 48 hours is identified as *C. albicans*. Colonies are negative for UV fluorescence at 48 hours.

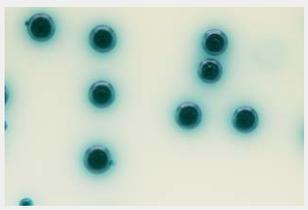
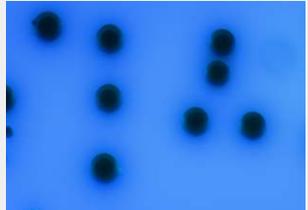
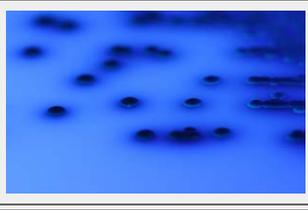
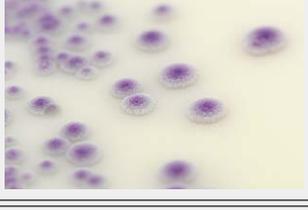
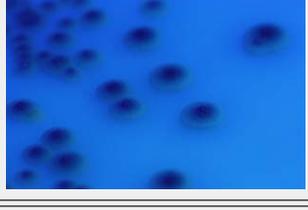
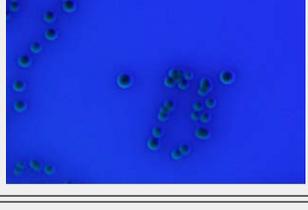
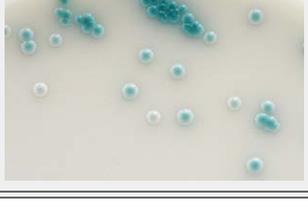
A medium size, smooth, medium blue to dark metallic blue colored colony, with a blue halo, at 48 hours is identified as *C. tropicalis*. Colonies are negative for UV fluorescence at 48 hours.

A large, flat, spreading, often rough or crenated, pink to medium pink colored colony is identified as *C. krusei*. Colonies are negative for UV fluorescence at 48 hours.

A medium size, smooth, pink colored colony, often with a darker mauve center, is presumptively identified as *C. glabrata*. A Rapid Trehalose test is needed (see "Limitations" below). Colonies are negative for UV fluorescence at 48 hours.

A medium size, smooth, white with teal to teal-green “bullseye” center at 48-72 hours is identified as *C. auris*. Colonies are positive for UV fluorescence at 48-72 hours.

Other yeast are generally small, white to pink colored colonies. Colonies are negative for UV fluorescence at 48 hours.

Organism	48hr Reaction	48hr Appearance	48hr Fluorescence
<i>Candida albicans</i>	Medium, smooth, green to dark metallic green colonies; (-) Fluorescence		
<i>Candida tropicalis</i>	Medium, smooth, medium blue to dark metallic blue colonies, with a blue halo; (-) Fluorescence		
<i>Candida krusei</i>	Large, flat, spreading, often rough or crenated, pink to medium pink colored colonies; (-) Fluorescence		
<i>Candida glabrata</i> *	Medium, smooth, pink colonies, often with a darker mauve center; (-) Fluorescence		
<i>Candida auris</i> **	Medium, smooth white colonies with a teal to teal-green "bullseye" center; (+) Fluorescence		

* A Rapid Trehalose test is needed (see "Limitations" below) to confirm *C. glabrata*.

** Full color development may take up to 72 hours for slow growing strains or for strains from mixed specimens.

LIMITATIONS

Some strains of *C. auris* may show a delayed color reaction and UV fluorescence in mixed specimens. This may also occur when the initial inoculum is low. In instances where delayed growth of *C. auris* is suspected, it is recommended to extend the incubation period to at least 72 hours. In addition, some strains of *C. auris* recovered in the presences of other yeast, such as *C. glabrata* or *C. krusei*, may exhibit a uniform teal to teal-green coloration instead of the characteristic "bullseye" colony center. In these instances, and as the *C. auris* colony color darkens over time, fluorescence is no longer detected. Therefore, incubation to 72 hours is only recommended when colonies show no characteristic color development at the 48 hour read time.

Other rare *Candida* species, as well as non-clinical yeast species, may exhibit fluorescence under UV light, but will not exhibit the characteristic white colonies with teal to teal-green "bullseye" centers at 48 to 72 hours indicative of *C. auris*.

C. haemulonii, which has been commonly misidentified for *C. auris*, exhibits fluorescence under UV light but produces

white colonies at 48 hours. *C. haemulonii* will eventually produce light pink or mauve colonies when incubated longer than 48 hours. Colonies will be positive for UV fluorescence.

Colonies of *C. glabrata* may show positive UV fluorescence at 24 hours, but the reaction will disappear by 48 hours.

Candida spp., other than *C. glabrata*, may present white to pink colored colonies on HardyCHROM™ *Candida* + *auris* which is why *C. glabrata* must be confirmed using Trehalose assimilation. Refer to the Rapid Trehalose Broth ([Cat. no. Z205](#)) or GlabrataQuick™ ([Cat. no. Z298](#)) technical information sheet for additional information concerning the definitive identification of *C. glabrata*. Colonies that are 24 hours old may be used to test for rapid Trehalose assimilation.

Isolates of *C. dubliniensis* will grow on this medium and will produce colors similar to or slightly different from *C. albicans* on primary isolation. The color variation will be lost upon subculture, so additional testing may be required to differentiate the two species. Both *C. dubliniensis* and *C. albicans* are germ tube positive, but *C. dubliniensis* can generally be differentiated from *C. albicans* by susceptibility to fluconazole, demonstration of TTC reduction (i.e. production of red colonies on media containing 2,3,5-triphenyl-tetrazolium), and poor to no growth at 45°C.⁽¹³⁾

This product is not intended for the isolation and identification of *Cryptococcus* spp.

Color-blind individuals may encounter difficulty in distinguishing the color differences on HardyCHROM™ *Candida* + *auris*.

It is recommended that biochemical, immunological, molecular, or mass spectrometry testing be performed on colonies from pure culture for complete identification.

Refer to the document "[Limitations of Procedures and Warranty](#)" for more information.

MATERIALS REQUIRED BUT NOT PROVIDED

Standard microbiological supplies and equipment such as loops, swabs, applicator sticks, UV lamp ([Cat. no. UVL56](#)), Rapid Trehalose Broth ([Cat. no. Z205](#)), GlabrataQuick™ ([Cat. no. Z298](#)), other culture media, incinerators, incubators, etc., as well as serological and biochemical reagents, are not provided.

QUALITY CONTROL

Hardy Diagnostics tests each lot of commercially manufactured media using appropriate quality control microorganisms and quality specifications as outlined on the Certificates of Analysis (CofA). The following organisms are routinely used for testing at Hardy Diagnostics:

Test Organisms	Inoculation Method*	Incubation			Results
		Time**	Temperature	Atmosphere	
<i>Candida albicans</i> ATCC® 10231	B	24-48hr	35°C	Aerobic	Growth; smooth, emerald green to dark metallic green colonies; negative UV fluorescence
<i>Candida tropicalis</i> ATCC® 750	B	24-48hr	35°C	Aerobic	Growth; smooth, medium blue to dark metallic blue colonies with a blue halo; negative UV fluorescence
<i>Candida krusei</i> ATCC® 14243	B	24-48hr	35°C	Aerobic	Growth; flat, pink to medium pink, large spreading, rough, crenated colonies;

					negative UV fluorescence
<i>Candida glabrata</i> ATCC® 66032	B	24-48hr	35°C	Aerobic	Growth; smooth pink colonies, often with a darker mauve center; negative UV fluorescence***
<i>Candida auris</i> CDC B11903	B	24-48hr	35°C	Aerobic	Growth; smooth, white colonies with teal to teal-green "bullseye" center; positive UV fluorescence
<i>Escherichia coli</i> ATCC® 25922	B	24hr	35°C	Aerobic	Partial to complete inhibition; negative UV fluorescence

* Refer to the document "[Inoculation Procedures for Media QC](#)" for more information.

** QC incubation period based on inoculum of pure isolates.

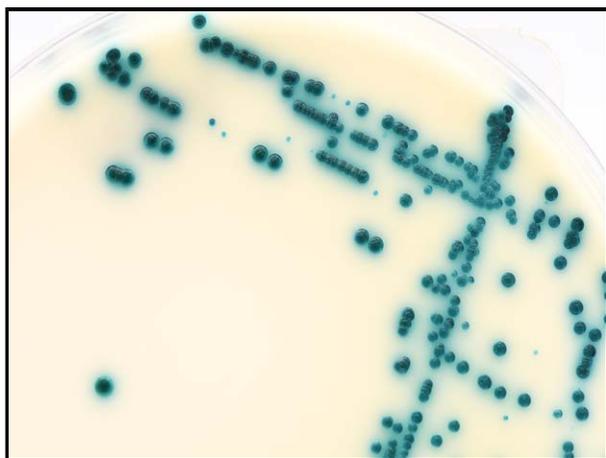
*** Fluorescence detected at 24hrs and disappears by 48hrs.

USER QUALITY CONTROL

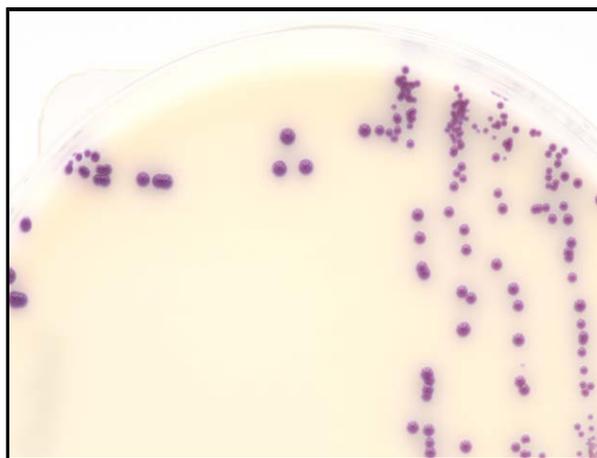
End users of commercially prepared culture media should perform QC testing in accordance with applicable government regulatory agencies, and in compliance with accreditation requirements. Hardy Diagnostics recommends end users check for signs of contamination and deterioration and, if dictated by laboratory quality control procedures or regulation, perform quality control testing to demonstrate growth or a positive reaction and to demonstrate inhibition or a negative reaction, if applicable. Hardy Diagnostics quality control testing is documented on the certificates of analysis (CofA) available from Hardy Diagnostics [Certificates of Analysis](#) website. In addition, refer to the following document "[Finished Product Quality Control Procedures](#)," for more information on QC or see reference(s) for more specific information.

PHYSICAL APPEARANCE

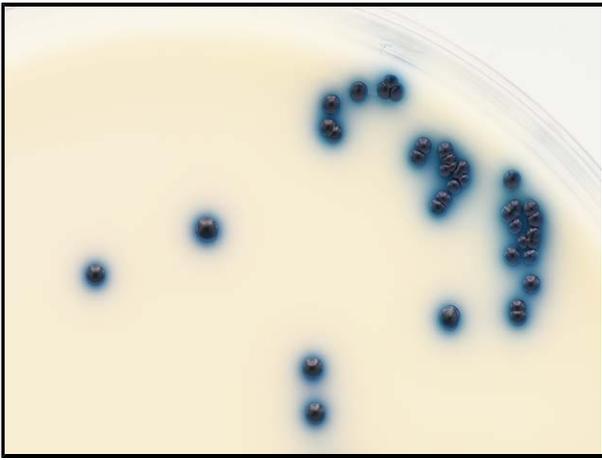
HardyCHROM™ *Candida + auris* should appear opaque and white in color.



Candida albicans (ATCC® 10231) colonies growing on HardyCHROM™ *Candida + auris* (Cat. no. G343). Incubated aerobically for 48 hours at 35°C.



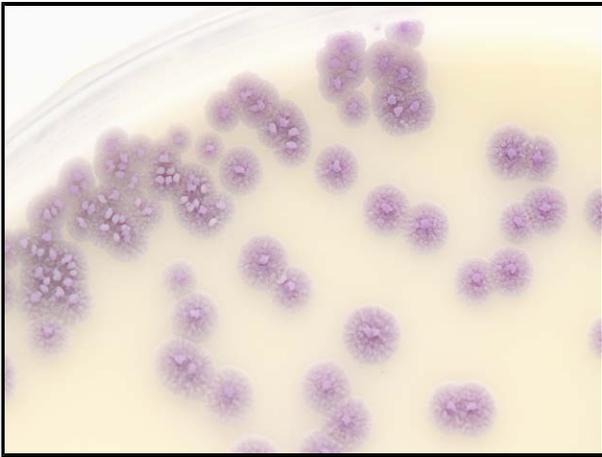
Candida glabrata (ATCC® 66032) colonies growing on HardyCHROM™ *Candida + auris* (Cat. no. G343). Incubated aerobically for 48 hours at 35°C.



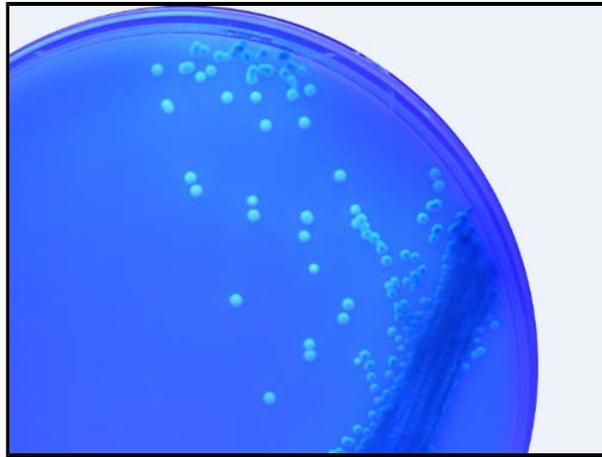
Candida tropicalis (ATCC® 750) colonies growing on HardyCHROM™ Candida + auris (Cat. no. G343). Incubated aerobically for 48 hours at 35°C.



Candida auris (CDC B11903) colonies growing on HardyCHROM™ Candida + auris (Cat. no. G343). Incubated aerobically for 48 hours at 35°C.



Candida krusei (ATCC® 14243) colonies growing on HardyCHROM™ Candida + auris (Cat. no. G343). Incubated aerobically for 48 hours at 35°C.



Candida auris (CDC B11903) colonies growing on HardyCHROM™ Candida + auris (Cat. no. G343) showing positive UV fluorescence. Incubated aerobically for 48 hours at 35°C.

REFERENCES

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13. Giammanco GM, G. Pizzo, S. Pecorella, S. Distefano, V. Pecoraro, M.E. Milici. April 2002. Identification of *Candida dubliniensis* among oral yeast isolates from an Italian population of human immunodeficiency virus-infected (HIV+) subjects. *Oral Microbiol Immunol*. 17(2):89-94.

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IFU-000801[B]



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